

VACCINE DEVELOPMENT USING LIVE *SALMONELLA ENTERICA* SV  
TYPHIMURIUM STRAINS DELETED IN FLAGELLIN -ENCODING GENES USING  
LAMBDA RED RECOMBINATION APPROACH

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*Salmonella* flagellins are potential carriers of vaccine antigens. Nonetheless, this system requires a flagellin-deficient strain that can produce recombinant flagellins fused to heterologous epitopes. *S. Typhimurium* has two non-allelic flagellin genes, *fliC* and *fljB*, which encode antigenically distinct proteins and the *fljA* gene which downregulates the *fliC* expression. Thus, the objective of the present study was the construction of *S. Typhimurium* strains deleted in the different flagellar genes. A non-flagellated strain was constructed using a site-specific recombination with a linear DNA fragment using the Red recombination system. Linear DNA fragments were amplified by PCR with downstream and upstream regions of the genes to be deleted (*fliC*, *fljBA*) and an antibiotic resistance marker. The site-specific recombination was obtained after transformation of a *S. Typhimurium* strain carrying a temperature-sensitive replication plasmid encoding the expression of the  $\lambda$  Red. The deletions were confirmed by PCR, motility tests and immunoblots with specific anti-flagellin antibody. This technique allowed the selection of stable *S. Typhimurium* mutants unable to express one or both flagellar antigens. The availability of strains defective in flagellin expression represents an important tool for the development of attenuated *Salmonella* strains that express recombinant flagellins fused to heterologous epitopes.

Supported by: FAPESP